**Mycobacterium avium** subsp. *paratuberculosis* Strains Isolated from Crohn’s Disease Patients and Animal Species Exhibit Similar Polymorphic Locus Patterns

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Crohn’s disease, a chronic inflammation of the intestine, results in substantial morbidity and medical costs in the United States (3). The etiology of Crohn’s disease is complex and appears to be associated with multiple factors for its pathological presentation (9; http://www.foodstandards.gov.uk/multimedia/pdfs/mapcrohnreport.pdf; http://gpvec.unl.edu/avc/IssuePapers/out38_en.pdf). The most prevalent theory of a microbial etiology of Crohn’s disease is infection with *Mycobacterium avium* subsp. *paratuberculosis*, the cause of Johnne’s disease in animals (8, 18; http://books.nap.edu/openbook/0309086116/html/index.html). Although several studies have associated *M. avium* subsp. *paratuberculosis* with a proportion of Crohn’s disease cases (5, 19), the evidence for a causal link remains controversial.

The primary objective of this report was to study the clonal distribution and degree of diversity of *M. avium* subsp. *paratuberculosis* isolated from humans with Crohn’s disease. Short sequence repeats (SSRs) have been successfully used as markers to understand the clonality and distribution of subtypes in several bacterial species (1, 7, 11). Preliminary SSR analysis of *M. avium* subsp. *paratuberculosis* isolates derived from a variety of hosts, disease types, and geographic localities suggests an association between allele types and host species (2, 14). Knowledge of strain specificity for the human host and genotypic proximity to isolates responsible for animal disease will be a critical first step in establishing a causal role for *M. avium* subsp. *paratuberculosis* in Crohn’s disease.

The human *M. avium* subsp. *paratuberculosis* isolates analyzed herein included those cultured from breast milk (16) and intestinal tissues (S. A. Naser et al., unpublished data; ATCC 43015, ATCC 43544, ATCC 43545, and ATCC 49164) of patients diagnosed with Crohn’s disease. The animal *M. avium* subsp. *paratuberculosis* isolates used included ATCC 700553, ATCC 19851, *M. avium* subsp. *paratuberculosis* K10, and those obtained from several animal host species with Johnne’s disease (Table 1) from different geographic localities (Fig. 1) and represented the extent of genotypic diversity determined by multiplex PCR of IS900 loci (15). *M. avium* subsp. *paratuberculosis* isolates from animals and birds in close proximity to cattle herds with multiple confirmed cases of Johnne’s disease were also analyzed (Table 1). All isolates were characterized by using well-defined molecular markers (14, 15) to confirm identity.

Results of a recently described multilocus SSR analysis (2) for *M. avium* subsp. *paratuberculosis* strain differentiation indicated that mononucleotide G repeat (GenBank accession no. AAK46234) and trinucleotide GGT repeat loci (GenBank accession no. CAB6859) were most discriminatory of the 11 SSRs analyzed. Hence, these loci were selected for fingerprinting strains in this study as previously described (2, 14). All chromatograms were manually edited and aligned with EditSeq and MegAlign, respectively (DNASTAR, Madison, Wis.). Alleles were identified by the number of G and GGT residues (Table 1). An allele containing a polymorphism within the repeat region was indicated by the letter *p*. Cluster analysis was performed with the molecular evolutionary genetic analysis program (version 2.1; www.megasoftware.net) by the neighboring method (12). The distance matrix for input into the molecular evolutionary genetic analysis program was created from the allele data with ETDIV and ETMEGA (http://foodsafemsu.edu/whittam/programs/). Simpson’s diversity index was calculated with the equation $1 - \Sigma(\text{allele frequency})^2$ (10).

Of the 94 *M. avium* subsp. *paratuberculosis* isolates analyzed, 92 had a detectable G repeat amplicon (approximately 400 bp) while all 94 had a detectable GGT repeat product (approximately 425 bp). Cluster analysis divided the isolates into three distinct clades and a total of 13 distinct alleles. Cattle ($n = 28$, including ATCC strains) and goat ($n = 20$) isolates were classified into nine and five alleles, respectively (Fig. 1). The sheep isolates ($n = 17$) were classified into eight alleles, three of which formed a distinct clade. Two sheep strain-specific nucleotide polymorphisms (GGGGGGG → GGGGGGG and GGTGGTGGT → AGTGGTGGT) [the nucleotides in ques-
tion are in italics) were identified within the repeat regions. Isolates with either of these polymorphic alleles had several additional substitutions throughout the sequenced region (GenBank accession numbers are in the footnotes to Table 1).

Two distinct alleles were identified among the 11 human isolates (including ATCC strains), and each clustered with cattle, sheep, and goat isolates. Isolates derived from free-ranging or feral nonruminant host species (n/H11005 18) were dispersed evenly in clades A and C (Fig. 1). Simpson’s diversity index for the analysis was 0.78 (D = 0.217), indicative of a strain discrimination capability much higher than that of other markers or methods reported to date (4, 15, 17).

While the number of alleles identified for cattle and goat isolates was greater than those in sheep isolates, 52% of the cattle and goat strains carried a single SSR genotype (7g-4ggt). This suggests a more recent association of *M. avium* subsp. *paratuberculosis* with cattle and goats compared to sheep. The presence of nucleotide polymorphisms, exclusive to sheep isolates, further supports the contention that sheep strains may be ancestral. This speculation is consistent with the data from a recently published study that indicated that the sheep strains of *M. avium* subsp. *paratuberculosis* were an evolutionary intermediate between the cattle strain of *M. avium* subsp. *paratuberculosis* and *M. avium* (6).

The restricted allelic variation identified within the human *M. avium* subsp. *paratuberculosis* strains analyzed may be in-

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**TABLE 1. SSR analysis results by host species and targets**

<table>
<thead>
<tr>
<th>Host</th>
<th>No. of <em>M. avium</em> subsp. <em>paratuberculosis</em> isolates</th>
<th>G repeat allele(s)</th>
<th>GGT repeat allele(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cow</td>
<td>28b</td>
<td>7, 8, 9, 10, 12, 14, 15</td>
<td>4, 5</td>
</tr>
<tr>
<td>Sheep</td>
<td>17</td>
<td>7, p7, 10, 14, 15</td>
<td>4, 5</td>
</tr>
<tr>
<td>Goat</td>
<td>20</td>
<td>7, 10, 11, 12, 15</td>
<td>4, 5</td>
</tr>
<tr>
<td>Deer</td>
<td>2</td>
<td>7</td>
<td>4</td>
</tr>
<tr>
<td>Human</td>
<td>11h</td>
<td>7</td>
<td>4, 5</td>
</tr>
<tr>
<td>Mouse</td>
<td>1</td>
<td>&gt;15</td>
<td>5</td>
</tr>
<tr>
<td>Raccoon</td>
<td>5</td>
<td>7, 14</td>
<td>5</td>
</tr>
<tr>
<td>Cat</td>
<td>1</td>
<td>14</td>
<td>5</td>
</tr>
<tr>
<td>Starling</td>
<td>7</td>
<td>7, 12, 13, 14, 15</td>
<td>5, 6</td>
</tr>
<tr>
<td>Shrew</td>
<td>1</td>
<td>15</td>
<td>5</td>
</tr>
<tr>
<td>Armadillo</td>
<td>1</td>
<td>7</td>
<td>4</td>
</tr>
</tbody>
</table>

* GenBank accession no. AY587703, AY587706, AY587707, AY587712, AY587714, AY587715, AY587717, AY587724, and AY587728.

b Includes 25 clinical strains, 2 animal ATCC strains, and K10.

c GenBank accession no. AY587699, AY587704, AY587708, AY587716, AY587718, AY587719, AY587720, AY587721, AY587725, and AY587729.

d Polymorphic 7-G repeats.

e Polymorphic 3-GGT repeats.

f GenBank accession no. AY587705, AY587709, AY587711, AY587713, AY587710, AY587726, and AY587730.

GenBank accession no. AY587700, AY587727, AY587701, AY587722, AY587702, and AY587723.

h Includes seven clinical strains and four human ATCC strains.

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**FIG. 1. Dendrogram showing the distribution of strains by the number of G and GGT repeats.** Shown are the number of G repeats followed by the number of GGT repeats and the host species. Host species not followed by a number indicate one isolate. The geographic locations (U.S. postal abbreviations) are in parentheses. A superscript following a geographic location indicates the number of isolates from that location. Also shown at the clade origins are the pairwise distances generated by the neighbor-joining model. unk, unknown.
indicative of the ability of a few animal genotypes to be associated with the pathobiology of Crohn’s disease (Fig. 2). The precedent for this line of thought lies in the acknowledgment that Johne’s disease in sheep is mostly caused by a distinct group of *M. avium* subsp. *paratuberculosis* strains. However, the presence of the same two alleles in 52% of the animal isolates is suggestive of strain sharing and interspecies transmission. Since the human *M. avium* subsp. *paratuberculosis* strains analyzed in this study (other than ATCC strains) were isolated from patients residing in Florida, the probability that they were exposed to a genetically and geographically restricted set of strains certainly exists. However, identification of the same alleles in ATCC strains obtained from diverse localities and the presence of disparate alleles in animal isolates (*n* = 4) from Florida strengthen the possibility that a limited set of strains are associated with Crohn’s disease. This inference is also reflected in the presence of several nonsynonymous single-nucleotide mutations unique to human *M. avium* subsp. *paratuberculosis* strains (X. Zhu and S. Sreevatsan, unpublished data). On the other hand, it is possible that the human *M. avium* subsp. *paratuberculosis* isolates from Florida represent a much broader geographic area than just the peninsula as many people retire to Florida from other parts of the country.

Isolation of *M. avium* subsp. *paratuberculosis* from tissues of other nonruminant animal species (cat, raccoon, shrew, armadillo, and rat) and birds found in the vicinity of infected farm animals suggests that *M. avium* subsp. *paratuberculosis* is able to infect and colonize nonruminants. Identification of common strain types between these animals and farm ruminants indicates strain sharing. Although we identified a greater representation of the 7g-5gt allele in the raccoon isolates, there was no evidence for an association between other nonruminant host species and a particular *M. avium* subsp. *paratuberculosis* allele such as that observed for human isolates.

It has been suggested that both genetic susceptibility (e.g., *NOD2/CARD15* polymorphisms) (13) and exposure to *M. avium* subsp. *paratuberculosis* are necessary for manifestation of Crohn’s disease. The existence of possible modes of exposure and/or transmission has been indicated by documentation of the presence of *M. avium* subsp. *paratuberculosis* in retail pasteurized milk and its long-term survival in soil and water (http://gpvec.unl.edu/avc/IssuePapers/out38_en.pdf). However, the prevalence of Crohn’s disease is low despite the suggested widespread exposure of people to *M. avium* subsp. *paratuberculosis* and isolates from humans do not reflect the strain diversity seen in animal *M. avium* subsp. *paratuberculosis* isolates. This further supports the idea that a few distinct *M. avium* subsp. *paratuberculosis* genotypes are associated with the pathobiology of Crohn’s disease. The present study presents evidence for the existence of both human disease-associated genotypes and strain sharing with animals. We were unable to establish if the patients carried the *NOD2/CARD15* mutation. Larger studies on *M. avium* subsp. *paratuberculosis* strains isolated from well-defined Crohn’s disease patient and healthy human populations from diverse geographic locations and time points are warranted to fully elucidate the association of specific genotypes of *M. avium* subsp. *paratuberculosis* with Crohn’s disease.

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REFERENCES


